KMCT
COLLEGE OF ALLIED HEALTH SCIENCES
Affiliated to Kerala University of Health Sciences and approved by Govt.of Kerala

TO WHOMSOEVER IT MAY CONCERN

This is to certify that the information in the attached documents is verified by me and is true to the best of my knowledge.

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INNOVATION AND ENTREPRENEURSHIP DEVELOPMENT CENTRE

4.2.3 Detailed report of activities in herbal garden

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HERBAL GARDEN TRIP - 2018 -2019

On 23/11/2018 a group of 93 students from KMCT CAHS along with three staff members embarked on a fruitful visit to a herbal garden. The visit aimed to provide students with hands-on experience and practical knowledge in the field of herbal medicine. Throughout the day, the students actively engaged in activities such as identifying and collecting plant samples, observing plant growth and characteristics, studying plant properties and uses, and learning about the extraction and preparation of herbal remedies. The staff members guided and facilitated the activities, ensuring a valuable and enriching experience for all participants. The visit concluded with discussions and reflections, allowing the students to share their insights and deepen their understanding of herbal medicine. The activity report highlights the successful participation of 92 students and the guidance provided by three staff members during the herbal garden visit, contributing to the students' overall learning and growth in the field of paramedical studies.

Staffs Accompanied: Mrs. Safa, Mrs. Dheeshma, Ms.Amina

ACTIVITY REPORT: TRAINING SESSION ON BIOMEDICAL WASTE MANAGEMENT

VENUE : KMCT CAHS AUDITORIUM

DATE : 18/03/2019

SPEAKER : Mrs Safeera, Associate Professor, Department of MLT, KMCT CAHS

SUMMARY: Mrs. Safeera from the Department of MLT conducted a useful session on Biomedical Waste Management, specifically targeting non-teaching staff such as cleaning and laboratory staff. The session likely focused on providing essential knowledge and guidelines for the proper disposal of biomedical waste. By addressing concerns and discussing doubts related to waste management, Her session aimed to ensure that the cleaning and laboratory staff have a clear understanding of the protocols and procedures for handling biomedical waste. This initiative contributes to a safer and more compliant environment within the institution.

BROCHURE ATTACHED

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HERBAL GARDEN- FIELD TRIP : 2019 - 2020

Date: 15/10/2019

Activities conducted

The identification of numerous herbs and plants that are frequently used in traditional medicine was presented to the students. They also observed these medicinal plants' growth patterns, leaf kinds, colors, and other distinguishing features. Through the investigation of plant properties and their uses, they were allowed to learn more about the therapeutic qualities and possible benefits of particular herbs, including the active ingredients they contain. Additionally, the students received lessons on various extraction processes and procedures used in the process of preparing herbal treatments. Additionally, they understood the usefulness of herbal remedies in a range of medical contexts, including topical applications and oral administration.

Participants

Faculties :Mrs.Binisha. M M, Ms. Dinu, M, Mr. Tajo

Students Participants : 89

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HERBAL GARDEN VISIT: 2020 - 2021

Introduction:

This activity report provides an overview of the Allied Students' visit to the Herbal Garden on 27/07/2021

Activities Conducted:

During the visit to the Herbal Garden, the following activities were conducted:

- The students received a guided tour of the Herbal Garden, where they learned about various medicinal plants, their uses, and cultivation techniques.
- During interactive sessions, students engaged with garden experts who shared their knowledge and answered questions about herbal plants.
- They were encouraged to identify different herbal plants in the garden, utilizing the information provided during the tour.
- A tasting session allowed the students to taste various herbal teas made from the garden's plants.
- Additionally, a workshop on making herbal remedies was conducted, enabling students to learn how to prepare simple herbal remedies for common ailments.

Number of students Participated : 88

Faculties : Mrs. Kashmeera P K, Ms. Balkees, Mrs. Chandini Kunjumon

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HERBAL GARDEN VISIT- 2021-2022

Date: 18/04/2022
Location: Manassery
Participants: 90 students + 4 Staffs
Staffs - Dr. Manju Suresh, Mrs. Jesna, Mrs. Anusruthi, Ms. Nimya

Objective:

The objective of the visit to the herbal garden was to provide students with an interactive and educational experience, enabling them to learn about various herbs, their uses, and the importance of conservation.

Activities:

1. Introduction and Orientation:

- The visit began with an introduction to the herbal garden and its significance in promoting health and wellbeing.

- Students were briefed on the rules and regulations of the garden to ensure their safety and respect for the environment.

2. Guided Tour:

- A knowledgeable guide led the students on a comprehensive tour of the herbal garden, highlighting various herbs and their properties.

- Students were encouraged to ask questions and actively engage in the learning process.

The guide demonstrated how to prepare herbal remedies, teas, and infusions using the harvested herbs.

- Students had the opportunity to taste and experience the various flavors and aromas of the prepared herbal products.

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ACTIVITY REPORT: KMCT AYURVEDA COLLEGE HERBAL GARDEN VISIT (2022-2023)

Date: 20/05/2023
Participants: 88 students
Staff accompanied : Mrs. Binisha, Ms. Drisya, Mrs. Sifana, Ms. Ananya, Mr. Jithin John, Mr. Jayadev, and Mrs. VIjisha

Objective:

- The objective of the visit to the herbal garden was to provide students with an interactive and educational experience, enabling them to learn about various herbs, their uses, and the importance of conservation.

Activities Conducted:

- The students were taken on a guided tour of the KMCT Ayurveda Herbal Garden. They were shown various herbs, plants, and trees, and their medicinal properties were explained in detail.
- Experts in Ayurveda conducted interactive sessions with the students, answering their questions and providing additional information about the herbs and their applications.
- The students were given the chance to touch, smell, and feel the different herbs and plants. They were encouraged to identify them based on their characteristics and learn about their individual uses.
- A question and answer session was organized, where students could ask further queries and clarify any doubts they had about Ayurveda and its practices.

Outcomes:

- The visit provided the students with valuable knowledge about Ayurveda herbs, their benefits, and their applications in traditional medicine.
- The interactive sessions and hands-on experience helped raise awareness about the importance of Ayurveda in promoting holistic well-being.

5. Feedback:

- Overall, the students found the Ayurveda Herbal Garden visit to be informative and engaging.
- Some students expressed a desire for more in-depth information about specific herbs and their applications.

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4.2.3

DETAILED REPORT OF ACTIVITIES IN LABORATORIES

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The laboratory activities in accordance with the KUHS syllabus aim to offer students a practical and interactive learning experience that allows them to connect their theoretical knowledge with real-world applications. Here's a comprehensive overview of the activities organized by each department:

The combination of mandatory practical, clinical postings, outposts, structured timetables, subject-specific registers, and computer lab access, along with strict attendance requirements, ensures that students receive a well-rounded and practical education in line with the KUHS syllabus.

Mandatory Practical Sessions: Practical sessions are mandatory for all four years of the program. These practicals are an essential component of the curriculum and play a crucial role in reinforcing theoretical concepts.

Total Hours as per Syllabus: The syllabus clearly outlines the total hours required for each academic year. This ensures that students receive comprehensive practical training aligned with the curriculum.

Class-wise Timetable: Class-wise timetables are thoughtfully designed to allocate dedicated hours for practical sessions. These designated hours are exclusively reserved for hands-on learning, ensuring students receive adequate exposure to laboratory work.

Attendance Requirement: To be eligible for examinations, it is imperative for all students to maintain over 80% attendance in practical and clinical postings. This requirement ensures students actively participate in these essential learning experiences.

Subject-wise Practical Registers: Subject-specific practical registers are meticulously maintained for microbiology, pathology, and biochemistry. Additionally, entries are made in the academic portal 'Campus Medicine'. This combination of record-keeping methods helps in maintaining accurate and easily accessible records for academic and administrative purposes

Clinical Postings: In addition to laboratory work within the institution, students participate in clinical postings at a central laboratory. This exposure to real-world healthcare settings enhances their understanding and application of theoretical knowledge

Computer Lab Access: Students have access to a computer lab, which supplements their practical learning. This allows them to analyse and document their findings, enhancing their overall laboratory skills.

List of Students in Laboratory Activities: Below is a list of students who have actively participated in laboratory activities.

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syllabus of BSc. MLT clearly outlines the total hours required for each academic year.

Subject		Total Hours/subject/year			
		Theory	Practical	Clinical Lab Posting	Total
	First Year				
Paper-I	I A Anatomy	100	80	-	180
	I B Physiology	115	65	-	180
Paper-II	Biochemistry-I	130	230	-	360
Paper III	Basic Microbiology & Immunology	130	230	-	360
Paper IV	Basic Medical Laboratory Science & Haematology – I	130	230		360
	Second year				
Paper V	Biochemistry II	90	174	96	360
Paper VI	General Microbiology	90	174	96	360
Paper VII	Parasitology & Entomology	90	174	96	360
Paper VIII	Haematology-II& Clinical Pathology	90	174	96	360
	Third year				
Paper IX	Biochemistry III	100	200	150	450
Paper X	Bacteriology	100	200	150	450
Paper XI	Cytology and Transfusion technology	100	200	150	450
Paper XII	Computer Application, Research methodology, Biostatistics & Laboratory management	90	--	--	90
	Final year				
Paper XIII	Biochemistry IV	90	180	160	430
Paper XIV	Mycology, Virology and Applied Microbiology	90	180	160	430
Paper XV	Histotechnology and Cytogenetics	90	180	160	430
Paper XVI	Project	150			150
	**Training at reputed external Hospitals/ National Institutions				100
	Grand Total				**5860**

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Practicals under each course of BSc. MLT programme

SPECIFICS OF PRACTICAL ASSIGNMENTS FOR EACH SUBJECT IN ACCORDANCE WITH THE KUHS CURRICULUM

ANATOMY

- Demonstration of Veins, arteries and nerves in the hands and legs
- Demonstration of bones Identification of normal tissues Human skeleton parts demonstration
- Preparation of histology slides
- Microscopic demonstration and identification of histology slides as per theory
- Museum jars, Preparation & Demonstration.
- Visit an Anatomy museum

PHYSIOLOGY

- Haemoglobin estimation
- ESR determination
- RBC count
- WBC count
- Differential count
- PCV, Red cell indices
- Osmotic fragility test
- Bleeding time, Clotting time
- Blood grouping
- Measurement of Blood pressure in man

BIOCHEMISTRY

BIOCHEMISTRY I

- Measurements of liquids, Weighing solids
- Calibration of pipette and other volumetric glass wares
- Preparation of saturated solution and half saturated solutions
- Preparation of standard solutions, % solutions (V/V. W/V normal and molar solutions.
- Preparation of buffers: acetate ,phosphate and tris buffers and measurement of pH
- Cleaning of lab wares and laboratory utensils, preparation of cleaning fluids.
- Preparation of distilled and deionised water Preparation of anticoagulants and preservatives for specimen collection.
- Use and proper maintenance of -Analytical balance, Electronic balance, Centrifuge, Colorimeter, spectrophotometer, pH meter, Homogenizer, Desiccators
- Measurement of pH,
- preparation of buffers
- Titration of acids and bases,
- preparation of standard solution of Sodium hydroxide, Hydrochloric acid, sulphuric acid. Silver nitrate and Potassium permanganate solutions
- Reactions of carbohydrates, reactions of glucose, fructose, maltose, lactose, sucrose, dextrin, starch and glycogen
- Reactions of Amino acids,
- colour reactions of albumin, globulin, casein, gelatin and peptone
- Reactions of fatty acids and cholesterol Reactions of NPN substances (urea, uric acid, creatinine)

BIOCHEMISTRY II

- Estimation and standardization of Blood/Serum/Plasma constituents glucose, Urea, Total protein, Albumin, Cholesterol, Triglyceride, Phospholipids, total lipid - Uric Acid, Creatine, Creatinine, Ammonia, Non- protein nitrogen, Amino Acid Nitrogen 2
- Qualitative detection of normal and abnormal constituents of Urine

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- Quantitative analysis for Urine protein, Bence Johnes's protein, Reducing sugars and chloride in Urine, Urea, Creatinine, Uric Acid, Aminoacids, Ammonia, Keto acids in Urine.
- Estimation of sugar, protein and chloride from C.S.F., plural fluid, peritoneal fluid, amniotic fluid foam test
- Glucose tolerance test and GCT
- Estimation of Ketone bodies in blood and urine.
- Estimation of Glycosylated Haemoglobin
- Estimation of Vitamin A,C,E and Metabolites of Vitamins in Urine (B complex)
- Tests for inborn errors of Amino Acid metabolism in Urine.

BIOCHEMISTRY III

- Activity determination of Clinically important enzymes- Alkaline Phosphates , Acid phosphates, Alanine amino transferase , Aspartate aminotransferase, Amylase, Ceruloplasmin, LDH, CPK and G6 PD
- Testing and semi quantitative assesment of urobilinogen in urine estimation of urobilin in urine, Estimation of porphyrin and porphobilonogen
- Estimation of bilirubin - direct and total.
- Qualitative analysis of Urinary calculi.
- Estimation of Haemoglobin, myoglobin and abnormal haemoglobins- Hb electrophoresis
- Identification of substances by column chromatography, Thin layer chromatography, paper chromatography, amino acids (Amino gram) and sugars.
- Technique of paper electrophoresis, agar gel electrophoresis of serum proteins, Polyacrylamide gel electrophoresis of serum proteins &Lipoproteins
- Clearance tests - Creatinine and Urea clearance
- Technique of RIA (T3, T4 and TSH) and ELISA.

BIOCHEMISTRY IV

- Estimation of calcium, Inorganic phosphorus, magnesium, Iron and Copper, Sodium and Potassium by flame Photometry
- Diagnosis of diseases with clinical correlation and Biochemical analysis of blood and Urine.
- Determination of clearance-urea and creatinine
- Gastric juice analysis Titrable acidity Test for malabsorption studies, D-Xylose, Stool fat, Occult blood in faeces
- Blood gas analysis, pH, PO2, PCO2. Estimation of bicarbonates
- Estimation of hormone metabolites in Urine - 17-Ketosteroids,17-Ketogenic Steroids, Urinary oestriol, Urinary VMA, 5 HIAA
- Familiarization and usage of all types of auto analyser
- Plotting of quality control charts and calculation of standard deviation

MICROBIOLOGY

BASIC MICROBIOLOGY AND IMMUNOLOGY

- Introduction and demonstration of Laboratory Equipment used in Microbiology
- Cleaning of new and used glass wares for microbiological purposes.
- Students be familiar to use autoclave, hot air oven, water bath, steamer etc
- Demonstration of different types of physical methods of sterilization
- Sterilization of heat labile fluids, glass wares, liquids, plastic and other laboratory and hospital wares.
- Demonstration of different methods of disinfection
- Students should be familiar to use different types of filters and its decontamination
- Rideal - Walker test or chick - Martin test
- Test for minimum inhibitory concentration of at least 2 commercially available disinfectants, In use test
- Students should prepare the working dilutions of common disinfectant
- Decontamination of wastes and carcasses - method

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Students should be trained to work with light microscope
Study of the morphological characters of bacteria
Motility of microbes
Hanging drop examinations with motile bacteria, non - motile bacteria. Method by using semi solid medium
Preparation and examination of wet films* Direct microscopic examination of wet film
Preparation of smear, fixation and staining of bacterial smears and its quality control methods 1%
Simple staining methods* – Pure culture, mixed culture
Gram's staining and any one modification* – Pure culture - Mixed culture
AFB staining and its modification * – Normal smear, AFB positive smear
Fluorochrome staining and its demonstration
Special staining technique for the demonstration of bacterial capsule (any two methods)
Special staining technique for the demonstration of spores (any one method)
Special staining technique for the demonstration of Flagella (Any one method)
Special staining technique for the demonstration of volutin granules
Preparation of stains and reagents used for the above staining technique. Quality control testing for the stain

GENERAL MICROBIOLOGY

- Preparation and use of pH indicator solutions
- Preparation of Reagents used for pH adjustments
- Adjustments of pH for Acidic medium and alkaline medium by using Lovibond Comparator
- Cleaning and preparation of glassware for media preparation and sterilization
- Preparation of sterile Saline.
- Students should be familiar with preparing the commonly used laboratory media and also they should know its sterilization, Quality control and storage. Peptone water, Nutrient broth, Nutrient agar, Blood agar, Chocolate agar, R.C.M ,Alk.Peptone Water, Selenite F broth, MaCconkey agar, XLD, TCBSA, L.J.medium, Transport medium (anyone) and other Media routinely used for the isolation for medically important bacteria.
- Preparation and standardization of bacterial loop.
- Inoculation methods on plate media, liquid media and slope media
- Inoculation and isolation of pure and mixed bacterial culture
- Study of colony characters on different media.
- Viable count of bacteria from a culture. Preparation of standard opacity tubes.
- Aerobic and Anaerobic incubation technique
- Preparation, Sterilization, Quality control, Inoculation and use of Biochemical media and its reagents used in bacteriology.
- Preparation of Reagents like methyl red indicator, V.P.Reagent, Nitrate reagents, Ferric Chloride. ONPG, H2O2, Oxidase reagent, Kovac's reagent, Ehrlich'sreagent.
- Inoculation methods and Quality control in different Biochemical media.
- Anaerobic cultivation methods - Anaerobic jar - Other methods Quality control
- Students should visit an Animal house and observe the organization and management of animal houses and its stock. Also they should observe the management of animals on experiments, safe handling of laboratory animals like Rabbit, Rat, Mouse, Guinea Pig

PARASITOLOGY AND ENTOMOLOGY

- Identification of parasites of medical importance dealt in the theory
- Macroscopic and microscopic examination of stool for adult worms, ova, cysts, larvae
- Concentration techniques for intestinal parasites in stool
- Collection of blood and preparation of thin and thick smears
- Staining of blood smears for blood parasites
- Examination of blood smears for malaria and microfilaria and their identification

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BACTERIOLOGY

- Identification of medically important bacteria from pure culture
  Staphylococcus • Streptococcus • Meningococcus, Gonococcus • Escherichia , Klebsiella, Serretia, Proteus, Salmonella, Shigella • Pseudomonas, Acinetobacter • Vibrio • Haemophilus
- Students should be familiar with the collection, transportation and processing of all type of clinical specimens for the diagnosis of bacterial infections discussed in theory
- Different methods & interpretation of antibiotic sensitivity tests.
- Isolation and identification of Mycobacterium tuberculosis from clinical specimens. Preparation of smear, staining, culture and reporting, concentration technique for the diagnosis of tuberculosis
- Examination of specimens from patient for the diagnosis of leprosy. Bacterial Serology
- Antisera preparation and standardization for Widal test. Widal test technique and interpretations
- VDRL Test, RPR, TPHA
- ASO Test
- Brucella agglutination test

VIROLOGY

- Demonstration of different type of cell lines.
- Demonstration of egg inoculation,
- Demonstration of CPE, Inclusion bodies, Paul Bunnel test,
- Demonstration of Viral Haemagglutination test, Viral Haemagglutination inhibition test, Viral neutralization test,
- Demonstration of immunofluorescence technique; Electron microscopy

MYCOLOGY

- Study of growth characteristics, microscopic examination and identification of medically important fungi, collection, transportation and processing of specimens for mycological examination.
- Slide culture technique
- Germ tube test for yeast identification

SEROLOGY

- Rosewaaler test, Latex agglutination test.
- Antinuclear antibody tests.
- Detection of C-Reactive protein.
- Haemolysin production and titration ELISA Examination of water- methods of collection of water and processing Presumptive coliform count and confirmatory tests
- Membrane filtration methods Examination of milk and milk products
- Preparation media and reagents for the study of water, food, milk and air
- Methylene blue test or phosphatase test,colony count test
- Milk ring test, Turbidity test, whey agglutination test
- Examination of food and food products
- Collection of samples and its processing –Frozen food, canned food and preserved food
- PCR technique and its modification

PATHOLOGY

HAEMATOLOGY I AND BMLS

- Care and use of light microscope
- capillary and venous blood collection
- Preparation of anticoagulant bottle
- Preparation of Romanowsky staining solutions
- Preparation of diluting fluids for cell counts
- Preparation of thick and thin smears and their staining

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- Haemoglobin estimation - cyan methaemoglobin method and Sahli's method
- ESR determination
- RBC count
- WBC count
- Differential count
- Platelet count
- Absolute eosinophil count
- Reticulocyte count
- PCV, Red cell indices
- Osmotic fragility test

HAEMATOLOGY II AND CLINICAL PATHOLOGY

- Peripheral blood smear examination and reporting
- Haemoglobin electrophoresis
- Blood cell cytochemistry- Peroxidase, PAS, LAP, Esterase
- Perl's stain
- Osmotic fragility test
- Sickling test
- LE cell demonstration
- Bleeding Time, Clotting Time, PT and APTT, clot retraction test, fibrinolysis test •
- Hemoglobin electrophoresis of myeloma proteins.
- Familiarisation of automation in Haematology
- Urine analysis, pregnancy test
- Examination of feces, detection of occult blood in stool, faecal urobilinogen and faecal fat detection
- Semen analysis
- Examination of CSF
- Examination of body fluids
- Examination of sputum

CYTOLOGY AND TRANSFUSION TECHNOLOGY

- Preparation of fixatives used in cytology
- Papanicolaou staining, May Grunwald Giemsa stain
- Shorr stain
- Processing and staining of various fluids for cytological examination
- Examination of normal and inflammatory cervicalsmears.
- Demonstration of normal cytology of respiratory tract, urinary tract, CSF, effusions.
- Preparation of 5% red cell suspension.
- ABO Blood grouping – cell grouping and serum grouping
- Rh typing methods
- Du typing
- Preparation of IgG coated cells, Direct and Indirect Antiglobulin tests
- Antibody titration
- Secretory status
- Screening tests done in donors
- Collection and storage of blood in blood bank
- Separation of packed red cells, FFP and cryoprecipitate

HISTOTECHNOLOGY AND CYTOGENETICS

- Preparation of commonly used fixatives- Formalin, Bouin's, Zenker's, Carnoy's
- Automatic tissue processors
- Decalcification
- Embedding

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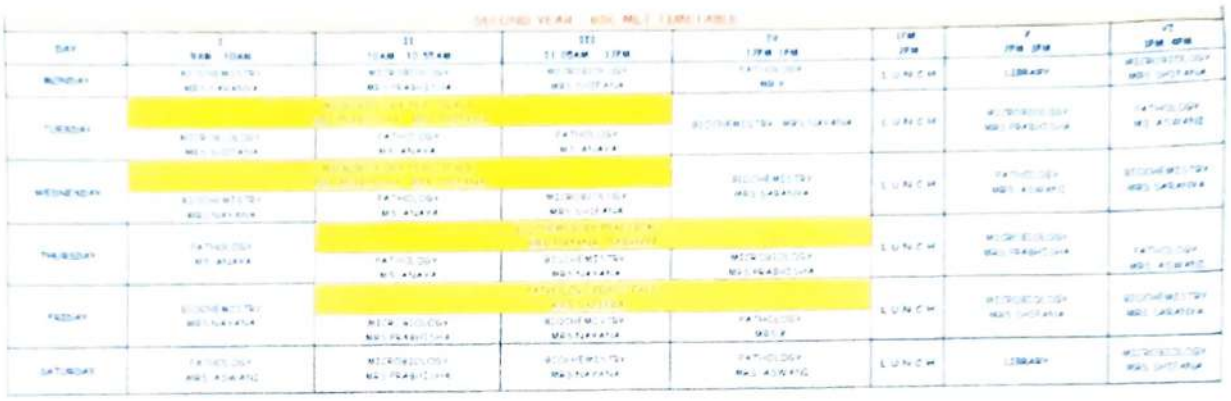
- **Class-wise timetables are designed to allocate dedicated hours for practical sessions**



FIRST YEAR BSC MLT TIMETABLE

DAY	I 9AM - 10AM	II 10AM - 10.55AM	III 11.05AM - 12PM	IV 12PM - 1PM	1PM - 2PM	V 2PM - 3PM	VI 3PM - 4PM
MONDAY	PATHOLOGY MLT ANATOMY	MICROBIOLOGY MLT PRACTICAL	BIOCHEMISTRY MLT THEORY	PATHOLOGY MLT	LUNCH	LIBRARY	LIBRARY
TUESDAY	BIOCHEMISTRY MLT PRACTICAL	ANATOMY MLT ANATOMY	PATHOLOGY MLT ANATOMY	LIBRARY	LUNCH	MICROBIOLOGY MLT PRACTICAL	BIOCHEMISTRY MLT THEORY
WEDNESDAY	PHYSIOLOGY AND BIOPHYSICS MLT ANATOMY AND PHYSIOLOGY			BIOCHEMISTRY MLT THEORY	LUNCH	PATHOLOGY MLT ANATOMY	PATHOLOGY MLT ANATOMY
THURSDAY	PATHOLOGY MLT ANATOMY	BIOCHEMISTRY MLT PRACTICAL AND THEORY		MICROBIOLOGY MLT PRACTICAL	LUNCH	MICROBIOLOGY MLT PRACTICAL	BIOCHEMISTRY MLT THEORY
FRIDAY	BIOCHEMISTRY MLT PRACTICAL	PHYSIOLOGY AND BIOPHYSICS MLT ANATOMY AND PHYSIOLOGY		PATHOLOGY MLT	LUNCH	MICROBIOLOGY MLT PRACTICAL	MICROBIOLOGY MLT PRACTICAL
SATURDAY	PATHOLOGY MLT ANATOMY	MICROBIOLOGY MLT PRACTICAL	BIOCHEMISTRY MLT THEORY	PATHOLOGY MLT ANATOMY	LUNCH	LIBRARY	LIBRARY

FIRST YEAR BSC MLT TIMETABLE

DAY	I 9AM - 10AM	II 10AM - 10.55AM	III 11.05AM - 12 PM	IV 12PM - 1PM	1PM - 2PM	V 2PM - 3PM	VI 3PM - 4PM
MONDAY	PATHOLOGY MLT ANATOMY	PHYSIOLOGY AND BIOPHYSICS MLT ANATOMY AND PHYSIOLOGY			LUNCH	PATHOLOGY MLT ANATOMY	LIBRARY
		MICROBIOLOGY MLT PRACTICAL	BIOCHEMISTRY MLT THEORY	PATHOLOGY MLT ANATOMY			
TUESDAY	BIOCHEMISTRY MLT PRACTICAL	ANATOMY MLT ANATOMY	PATHOLOGY MLT ANATOMY	LIBRARY	LUNCH	PATHOLOGY MLT ANATOMY	BIOCHEMISTRY MLT ANATOMY
WEDNESDAY	PHYSIOLOGY AND BIOPHYSICS MLT ANATOMY AND PHYSIOLOGY			BIOCHEMISTRY MLT THEORY	LUNCH	MICROBIOLOGY MLT PRACTICAL	PATHOLOGY MLT ANATOMY
	BIOCHEMISTRY MLT PRACTICAL	MICROBIOLOGY MLT PRACTICAL	MICROBIOLOGY MLT PRACTICAL				
THURSDAY	PHYSIOLOGY AND BIOPHYSICS MLT ANATOMY AND PHYSIOLOGY			BIOCHEMISTRY MLT THEORY	LUNCH	PATHOLOGY MLT ANATOMY	MICROBIOLOGY MLT PRACTICAL
	PATHOLOGY MLT ANATOMY	BIOCHEMISTRY MLT PRACTICAL	PATHOLOGY MLT ANATOMY				
FRIDAY	MICROBIOLOGY MLT PRACTICAL	PHYSIOLOGY MLT PRACTICAL	ANATOMY MLT ANATOMY	PATHOLOGY MLT PRACTICAL	LUNCH	PATHOLOGY MLT ANATOMY	MICROBIOLOGY MLT PRACTICAL
SATURDAY	MICROBIOLOGY MLT PRACTICAL	PATHOLOGY MLT PRACTICAL	ANATOMY MLT ANATOMY	ANATOMY MLT ANATOMY	LUNCH	MICROBIOLOGY MLT PRACTICAL	BIOCHEMISTRY MLT THEORY

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- **Class-wise timetables designed to allocate dedicated hours for practical sessions.**

KMCT COLLEGE OF ALLIED HEALTH SCIENCES – DEPT.OF MLT- TIMETABLE: 2022 – 23 ACADEMIC YEAR

FOURTH YEAR BSC MLT TIMETABLE

DAY	I 9AM - 10AM	II 10AM - 10.55AM	III 11.05AM - 12 PM	IV 12PM-1PM	1PM - 2PM	V 2PM - 3PM	VI 3PM - 4PM
MONDAY	MICROBIOLOGY PRACTICAL MRS PRABISHA, MRS AMRUTHA, MRS SARANYA			LIBRARY	LUNCH	BIOCHEMISTRY MRS DEESHMA	MICROBIOLOGY MRS AMRUTHA
TUESDAY	PATHOLOGY MS ANANYA	MICROBIOLOGY MRS AMRUTHA	PROJECT	PATHOLOGY PRACTICAL MRS SOUMYA, MS ANANYA, MRS DEESHMA	LUNCH	BIOCHEMISTRY PRACTICAL MRS DEESHMA, MRS SARANYA, MRS AMRUTHA	
WEDNESDAY	BIOCHEMISTRY MRS DEESHMA	MICROBIOLOGY MRS PRABISHA	PROJECT	MICROBIOLOGY PRACTICAL MRS PRABISHA, MRS AMRUTHA, MRS SARANYA	LUNCH	PATHOLOGY MRS SOUMYA	PATHOLOGY MS ANANYA
THURSDAY	MICROBIOLOGY MRS PRABISHA	PATHOLOGY MRS SOUMYA	BIOCHEMISTRY MRS SARANYA	LIBRARY	LUNCH	PATHOLOGY PRACTICAL MRS SOUMYA, MS ANANYA, MRS DEESHMA	
FRIDAY	BIOCHEMISTRY PRACTICAL MRS DEESHMA, MRS SARANYA, MRS AMRUTHA			PATHOLOGY MS ANANYA	LUNCH	BIOCHEMISTRY MRS SARANYA	PATHOLOGY MRS SOUMYA
SATURDAY	PATHOLOGY MRS SOUMYA	BIOCHEMISTRY MRS SARANYA	MICROBIOLOGY MRS AMRUTHA	BIOCHEMISTRY MRS DEESHMA	LUNCH	PATHOLOGY MS ANANYA	MICROBIOLOGY MRS PRABISHA

THIRD YEAR BSC MLT TIMETABLE

DAY	I 9AM - 10AM	II 10AM - 10.55AM	III 11.05AM - 12PM	IV 12PM-1PM	1PM - 2PM	V 2PM - 3PM	VI 3PM - 4PM
MONDAY	PATHOLOGY PRACTICAL MRS SOUMYA, MRS NAYANA, MRS SUSI			PATHOLOGY MRS SOUMYA	LUNCH	BIOCHEMISTRY MRS ANDORA	MICROBIOLOGY MRS SUSI
TUESDAY	BIOCHEMISTRY PRACTICAL MRS ANDORA, MRS V, MRS NAYANA			MICROBIOLOGY MRS AMRITHA	LUNCH	MICROBIOLOGY MRS SUSI	BIOCHEMISTRY MRS NAYANA
WEDNESDAY	RESEARCH	BIOCHEMISTRY MRS NAYANA	PATHOLOGY MRS V	BIOCHEMISTRY MRS ANDORA	LUNCH	LIBRARY	MICROBIOLOGY MRS AMRITHA
THURSDAY	MICROBIOLOGY PRACTICAL MRS SUSI, MRS ANDORA, MRS AMRITHA			RESEARCH	LUNCH	MICROBIOLOGY MRS SUSI	PATHOLOGY
FRIDAY	MICROBIOLOGY PRACTICAL MRS SUSI, MRS ANDORA, MRS AMRITHA			PATHOLOGY MRS SOUMYA	LUNCH	LIBRARY	MICROBIOLOGY MRS AMRITHA
SATURDAY	MICROBIOLOGY PRACTICAL MRS SUSI, MRS ANDORA, MRS AMRITHA			BIOCHEMISTRY MRS NAYANA	LUNCH	PATHOLOGY MRS SOUMYA	BIOCHEMISTRY MRS ANDORA

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FIRST YEAR BPT PRACTICAL HOUR DETAILS

FIRST YEAR BPT (DURATION 0 – 12 MONTHS)					
S No.	Subjects	Teaching hours			
		Hours/Week	Theory	Practical	Total
Main subjects: For University Examination					
01	Anatomy	8	160	160	320
02	Physiology	8	160	160	320
03	Biomechanics & Kinesiology	8	160	160	320
04	Psychology	2	80	-	80
05	Sociology	2	80	-	80
Subsidiary subjects: Not for University Examination					
06	Biochemistry	2	80	-	80
07	First Aid & Nursing	2	60	20	80
08	Nutrition	1	40		40
09	Orientation to Physiotherapy	1	40	-	40
10	Communicative English	1	40	-	40
11	Seminar	1	40	-	40
12	Total	36	910	530	1440

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SECOND YEAR BPT PRACTICAL HOUR DETAILS

SECOND YEAR BPT (DURATION 13 – 24 MONTHS)					
S No.	Subjects	Teaching hours			
		Hours/Week	Theory	Practical/Clinical	Total
Main subjects: For University Examination					
01	Electrotherapy	8	120	200	320
02	Exercise therapy	8	120	200	320
03	Pharmacology	2	80	-	80
04	Microbiology	2	80	-	80
05	Pathology	2	80	-	80
Subsidiary subjects: Not for University Examination					
06	Computer Science	2	40	40	80
07	Medical Instrumentation	2	60	20	80
08	Seminar	2	80	-	80
09	Supervised Clinical Observation	8	-	320	320
10	Total	36	660	780	1440

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THIRD YEAR BPT PRACTICAL HOUR DETAILS

THIRD YEAR BPT (DURATION 25 – 36 MONTHS)					
S No.	Subjects	Teaching hours			
		Hours/Week	Theory	Practical/Clinical	Total
Main subjects: For University Examination					
01	General Medicine & General Surgery	2	80	-	80
02	Physiotherapy in General Medicine & General Surgery	6	80	160	240
03	Cardio- Respiratory disorders & Surgery	2	80	-	80
04	Physiotherapy in Cardio-Respiratory disorders & Intensive Care management	6	80	160	240
05	Community Medicine	2	80	-	80
Subsidiary subjects: Not for University Examination					
06	Research methodology & Biostatistics	2	80	-	80
07	Ethics and Management	1	40	-	40
08	Seminar	1	40	-	40
09	Supervised Clinical Practice	14	-	560	560
10	Total	36	560	880	1440

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FOURTH YEAR BPT PRACTICAL HOUR DETAILS

FORTH YEAR BPT (DURATION 37– 48 MONTHS)					
S No.	Subjects	Teaching hours			
		Hours/Week	Theory	Practical/Clinical	Total
Main subjects: For University Examination					
01	Neurology & Neurosurgery	2	80	-	80
02	Physiotherapy in Neurology & Neurosurgery	6	80	160	240
03	Orthopedics & Sports Medicine	2	80	-	80
04	Physiotherapy in Orthopedics & Sports	6	80	160	240
05	Physiotherapy in Community Health & Project	6	80	160	240
Subsidiary subjects: Not for University Examination					
06	Supervised Clinical Practice	14	-	560	560
07	Total	36	400	1040	1440

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SPECIFICS OF PRACTICAL ASSIGNMENTS FOR EACH SUBJECT IN ACCORDANCE WITH THE KUHS CURRICULUM FOR BPT STUDENTS

ANATOMY

- Upper extremity Anatomy
- Lower extremity Anatomy
- Head & Spinal cord and Neck and Brain including surface Anatomy
- Thorax including surface anatomy, abdominal muscles, joints, Diaphragm
- Embryology- Histology-Elementary tissue including surface Anatomy
- Demonstration of the muscles of the whole body and organs in Thorax and Abdomen in a cadaver
- Demonstration of movements & discuss about the range of motion (ROM) in important joints. c. Surface marking of the lung, pleura, fissures and lobes of lungs, heart, liver, spleen and muscles.
- Kidney, cranial nerves, spinal nerves and important blood vessels.
- Identification of body prominences on inspection and by palpation especially of extremities.
- Points of palpation of muscles, tendons, bones, joints, ligaments, nerves and arteries

PHYSIOLOGY

- Haematology
- Study of Microscope and its uses
- Determination of blood groups
- Determination of bleeding time
- Determination of clotting time
- Determination of ESR
- Determination of PC V
- Examination of Radial pulse.
- Recording of blood pressure
- Examination of CVS
- Examination of Respiratory system
- Examination of sensory system
- Examination of Motor System
- Examination of reflexes
- Spirometry, Body Composition, Exercise testing.
- Ergometry, Artificial Respiration
- ECG, EEG

BIOMECHANICS AND KINESIOLOGY

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- Practical shall be conducted for various joint movements and analysis of the same.
- Demonstrations should also be given as how to analyze posture and gait.
- The student shall be taught and demonstrated to analysis for activities of daily living – ADL – (sitting to standing, throwing, lifting etc.)
- The student should be able to explain and demonstrate the movements occurring at the joints, the muscles involved, the movements or muscle action produced, and mention the axis and planes through which the movements occur.
- The demonstrations may be done on models or skeleton

ELECTROTHERAPY

- Demonstrate the technique for patient evaluation - receiving the patient and positioning the patient for treatment using electrotherapy.
- Collection of materials required for treatment using electrotherapy modalities and testing of the apparatus.
- Demonstrate placement of electrodes; for various electrotherapy modalities
- Electrical stimulation for the muscles supplied by the peripheral nerves
- Faradism under Pressure for UL and LL
- Plotting of SD curve with chronaxie and rheobase.
- Demonstrate FG test
- Application of Ultrasound for different regions and various methods of application
- Demonstrate treatment techniques using SWD. IRR and Microwave diathermy
- Demonstrate treatment method using IFT for various regions
- Calculation of dosage and technique of application of LASER
- Technique of application of Hydrocollator packs, cryotherapy, contrast bath, Wax therapy
- Demonstrate the treatment method using Whirl pool bath
- Winding up procedure after any electrotherapy treatment method.
- Demonstration of methods for basic maintenance and repair of all Electrotherapy Equipments.

EXERCISE THERAPY

- Demonstrate the technique of measuring using goniometry
- Demonstrate muscle strength using the principles and technique of MMT
- Demonstrate – Basic Asana, Pranayama and Meditation methods
- Demonstrate the PNF techniques
- Demonstrate exercises for training co-ordination – Frenkel's exercises
- Demonstrate the techniques of massage and Soft Tissue manipulations
- Demonstrate technique for functional re-education
- Assess and train for using walking aids
- Demonstrate mobilization of individual joint regions
- Demonstrate to use the technique of suspension therapy for mobilizing and strengthening joints and muscles
- Demonstrate the techniques for muscle stretching
- Assess and evaluate posture and gait

- Demonstrate to apply the technique of passive movements. Demonstrate various techniques of active movements
- Demonstrate techniques of strengthening muscles using resisted exercises
- Demonstrate techniques for measuring limb length and body circumference

PHYSIOTHERAPY IN GENERAL MEDICINE AND GENERAL SURGERY

- Bedside case presentations and case discussions in the wards
- Demonstration of application of general physiotherapeutic techniques on patients
- Participation under faculty guidance in management.
- Lab sessions consisting of evaluation and assessment methods on student models, treatment techniques and practice sessions

PHYSIOTHERAPY IN CARDIO-RESPIRATORY DISORDERS & INTENSIVE CARE MANAGEMENT

- Bedside case presentations and case discussions in the wards
- Demonstration of application of cardio-respiratory physiotherapeutic techniques on patients
- Participation under faculty guidance in management.
- Lab sessions consisting of evaluation and assessment methods on student models, treatment techniques and practice sessions

PHYSIOTHERAPY IN NEUROLOGY & NEUROSURGERY

- Bedside case presentations and case discussions in the wards
- Demonstration of application of Neuro Physiotherapeutic techniques on patients in Physiotherapy O.P.D
- Participation under faculty guidance in management.
- Lab sessions consisting of evaluation and assessment methods on student models, treatment techniques and practice sessions.

PHYSIOTHERAPY IN ORTHOPEDICS & SPORTS

- Bedside case presentations and case discussions in the wards
- Demonstration of application of Physiotherapeutic techniques on patients in Physiotherapy O.P.D
- Participation under faculty guidance in management.
- Lab sessions consisting of evaluation and assessment methods on student models,treatment techniques and practice sessions.

PHYSIOTHERAPY IN COMMUNITY HEALTH

- Bedside case presentations and case discussions in the wards
- Demonstration of application of Physiotherapeutic techniques on patients in Physiotherapy O.P.D
- Participation under faculty guidance in management.
- Lab sessions consisting of evaluation and assessment methods on student models,treatment techniques and practice sessions.

BSC OPTOMETRY

CLINICAL EXAMINATION OF VISUAL SYSTEMS

Sl. No.	Topic	No. of hours
1.	History taking,	4
2.	Visual acuity estimation	1
3.	Extraocular motility, Cover teat, Alternating cover test	2
4.	Hirschberg test, Modified Krimsky,	1
5.	Pupils Examination	1
6.	Maddox Rod,	1
7.	van Herrick,	1
8.	External examination of the eye, Lid Eversion	1
9.	Schirmer's, TBUT, tear meniscus level, NITBUT (keratometer),	2
10.	Color Vision	1
11.	Stereopsis,	1
12.	Confrontation test,	1
13.	Photostress test,	1
14.	Slitlamp biomicroscopy,	3
15.	Direct Ophthalmoscopy,	1
16.	Digital pressure, Schiotz Tonometry, Applanation Tonometry Gonioscopy	3
17.	ROPLAS	1
18.	Amsler test,	1
19.	Corneal Sensitivity, HVID	1
20.	Saccades and Pursuits	1
	Total no. hours	**29**

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1. OPTOMETRIC OPTICS

No.	Topic	No. of Lectures
1	Introduction – Light, Mirror, Reflection, Refraction and Absorption	1
2	Prisms – Definition, properties, Refraction through prisms, Thickness difference, Base-apex notation, uses, nomenclature and units, Sign Conventions, Fresnel's prisms, rotary prims	4
3	Lenses – Definition, units, terminology used to describe, form of lenses	2
4	Vertex distance and vertex power, Effectivity calculations	2
5	Lens shape, size and types i.e. spherical, cylindrical and Sphero-cylindrical	1
6	Transpositions – Simple, Toric and Spherical equivalent	1
7	Prismatic effect, centration, decent ration and Prentice rule, Prismatic effect of Plano-cylinder and Sphero-cylinder lenses	4
8	Spherometer & Sag formula, Edge thickness calculations	3
9	Magnification in high plus lenses, Minification in high minus lenses	1
10	Tilt induced power in spectacles	1
11	Aberration in Ophthalmic Lenses	1

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No.	Topic	No. of Lectures
1	Raw materials – History and General Outline, Manufacturing of Ophthalmic Blanks – Glass & Plastics, Terminology used in Lens Workshops, Surfacing process from Blanks to lenses	1 4
2	Definition & Materials (Glass, Plastics, Polycarbonate, Triology) types and Characteristics	4
3	Properties (Refractive index, specific gravity, UV cut off, impact resistance – include drop ball test, abbe value, Center thickness)	4
4	Best form of lenses & Safety standards for Ophthalmic lenses (FDA, ANSI, ISI, Others)	2
5	Design of High Powered Lenses Hi-index lenses, Calculation of Refractive index	2
6	Bifocal designs, their manufacturing & uses (Kryptok, Univis D, Executive, Invisible, Occupational)	6
7	Progressive Addition Lenses, modified near vision lenses (designs, advantages, limitations)	3
8	Lens enhancements (Scratch resistant coatings – spin/dip, Anti-reflection coating, UV coating, Hydrophobic coating, anti-static coating	4
9	Lens defects – Description and Detection	2
10	Glazing & edging (manual & automatic)	2
11	Special lenses • Lenticulars • Aspherics • Fresnel lenses & Prisms • Aniseikonic lenses • Photochromics • Polaroids • Tinted lenses – Tints, filters	6
12	History of Spectacles, manufacturing overview, Definition, parts & measurements	2
13	Classification of frames – Materials (cover in detail), Colours and Temple position (advantages & disadvantages, where to use)	4
14	Special purpose frames (sports, kids, reading)	1
	Total hours	42

3. VISUAL OPTICS

Topic	No. of hours
1.Refractive conditions - Emmetropia - Myopia - Hyperopia - Astigmatism - Accommodation - Presbyopia - Anisometropia and Aniseikonia - Aphakia and Pseudophakia	9 hours
2.Accommodation - Far and near points of accommodation - Correction of spherical ametropia - Axial versus refractive ametropia - Relationship between accommodation and convergence, AC / A ratio	6 hours
3.Objective refraction - Streak Retinoscopy only	3 Hours
4.Subjective Refraction - Review of subjective refractive methods - Cross cylinder methods for astigmatism, Astigmatic Fan Test - Difficulties in subjective and objective tests and their avoidance - Ocular refraction versus spectacle refraction - Ocular accommodation versus spectacle accommodation - Spectacle magnification and relative spectacle magnification - Retinal image blur; depth of focus and depth of field - Prescribing Prisms / Binocular Refraction	9 hours
Total no. of hours	**27 hours**

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5. EYE DISEASES

S. No.	Topics	No. of Lectures
1.	a) **ORBIT** ➢ Applied Anatomy ➢ Proptosis Classification,Causes,Investigations) ➢ Enophthalmos ➢ Developmental Anomalies ➢ (craniosynostosis, Craniofacial Dysostosis, Hypertelorism, Median facial cleft syndrome) ➢ Orbital Inflammations (Preseptal cellulites, Orbital cellulitis Orbital Periostitis, cavernous sinus Thrombosis) ➢ Grave's Ophthalmopathy ➢ Orbital tumors(Dermoids, capillary haemangioma, Optic nerve glioma) ➢ Orbital blowout fractures ➢ Orbital surgery (Orbitotomy) ➢ Orbital tumors ➢ Orbital trauma ➢ Approach to a patient with proptosis	10
2.	b) **LIDS** Applied Anatomy ➢ Congenital anomalies (Ptosis, Coloboma, Epicanthus, Distichiasis, Cryptophthalmos)	6

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	➢ 3.Oedema of the eyelids (Inflammatory, Solid, Passive edema) ➢ Inflammatory disorders (Blepharitis, External Hordeolum, Chalazion, ➢ Internal hordeolum,.Molluscum Contagiosum) ➢ Anomalies in the position of the lashes and Lid Margin (Trichiasis, Ectropion, Entropion, Symblepharon, Blepharophimosis, Lagophthalmos, Blepharospasm, Ptosis). ➢ Tumors (Papillomas, Xanthelasma, Haemangioma, Basal carcinoma, Squamous cell carcinoma, sebaceous gland melanoma)	
3.	c) **LACRIMAL SYSTEM** ➢ Applied Anatomy ➢ Tear Film ➢ The Dry Eye (Sjogren's Syndrome) ➢ The watering eye (Etiology, clinical evaluation) ➢ Dacryocystitis ➢ Swelling of the Lacrimal gland (Dacryoadenitis)	4
4.	d) **CONJUNCTIVA** ➢ Applied Anatomy ➢ Inflammations of conjunctiva (Infective conjunctivitis – bacterial, chlamydial, viral , Allergic conjunctivitis, Granulomatous conjunctivitis) ➢ Degenerative conditions ➢ (Pinguecula, Pterygium, Concretions) ➢ Symptomatic conditions ➢ (Hyperaemia, Chemosis, Ecchymosis, Xerosis, Discoloration) ➢ 5.Cysts and Tumors	4
5.	e) **CORNEA** ➢ Applied Anatomy and Physiology ➢ Congenital Anomalies (Megalocornea, Microcornea, Cornea plana, Congenital cloudy cornea) ➢ Inflammations of the cornea (Topographical classifications: Ulcerative keratitis and Non ulcerative ➢ Etiological classifications: Infective, Allergic, Trophic, Traumatic, Idiopathic) ➢ Degenerations (classifications, Arcus senilis, Vogt's white limbal girdle, Hassal-henle bodies, Lipoid Keratopathy, Band shaped keratopathy, Salzmann's nodular degeneration, Droplet keratopathy, Pellucid Marginal degeneration) ➢ Dystrophies (Reis Buckler dystrophy,Recurrent corneal erosion syndrome, Granualr dystrophy,Lattice dystrophy, Macular dystrophy, cornea guttata, Fuch's epithelial endothelial dystrophy, Congenital hereditary endothelial dystrophy)	12

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	➢ Keratoconus, Keratoglobus ➢ Corneal oedema, Corneal opacity, Corneal vascularisation ➢ Penetrating Keratoplasty	
6.	f) **UVEAL TRACT AND SCLERA** ➢ Applied Anatomy, ➢ Classification of uveitis ➢ Etiology ➢ Pathology ➢ Anterior Uveitis ➢ Posterior Uveitis ➢ Purulent Uveitis ➢ Endophthalmitis ➢ Panophthalmitis ➢ Pars Planitis ➢ Tumors of uveal tract(Melanoma) ➢ Episcleritis and scleritis ➢ Clinical examination of Uveitis and Scleritis	10
7.	Retina and Vitreous: ➢ Applied Anatomy ➢ Congenital and Developmental Disorders (Optic Disc: Coloboma, Drusen, Hypoplasia, Medullated nerve fibers; Persistent Hyaloid Artery) ➢ Inflammatory disorders (Retinitis : Acute purulent , Bacterial, Virus, mycotic) ➢ Retinal Vasculitis (Eales's) ➢ Retinal Artery Occlusion (Central retinal Artery occlusion) ➢ Retinal Vein occlusion (Ischaemic, Non Ischaemic , Branch retinal vein occlusion) ➢ Retinal degenerations : Retinitis Pigmentosa, Lattice degenerations ➢ Macular disorders: Solar retinopathy, central serous retinopathy, cystoid macular edema, Age related macular degeneration. ➢ Retinal Detachement: Rhegmatogenous, Tractional, Exudative) ➢ Retinablastoma	12

8.	Ocular Injuries Terminology : Closed globe injury (contusion, lamellar laceration) Open globe injury (rupture, laceration, penetrating injury, peforating injury) Mechanical injuries (Extraocular foreign body, blunt trauma, perforating injury, sympathetic ophthalmitis) - Non Mechanical Injuries (Chemical injuries, Thermal, Electrical, Radiational) - Clinical approach towards ocular injury patients	4
9.	Lens - Applied Anatomy and Physiology - Clinical examination - Classification of cataract - Congenital and Developmental cataract - Acquired (Senile, Traumatic, Complicated, Metabolic, Electric, Radiational, Toxic) - Morphological: Capsular, Subcapsular, Cortical, Supranuclear, Nuclear, Polar. - Management of cataract (Non surgical and surgical measures; preoperative evaluation, Types of surgeries,) - Complications of cataract surgery - Displacement of lens: Subluxation, Displacement - Lens coloboma, Lenticonus, Microsperophakia.	10
10.	Clinical Neuro-ophthalmology - Anatomy of visual pathway - Lesions of the visual pathway - Pupillary reflexes and abnormalities (Amaurotic light reflex, Efferent pathway defect, Wernicke's hemianopic pupil, Marcus gunn pupil. Argyll Robetson pupil, Adie's tonic pupil) - Optic neuritis, Anterior Ischemic optic neuropathy, Pappilloedema, optic atrophy - Cortical blindness - Malingering - Nystagmus - Clinical examination	12
11.	Glaucoma - Applied anatomy and physiology of anterior segment - Clinical Examination	10
	- Definitions and classification of glaucoma - Pathogenesis of glaucomatous ocular damage - Congenital glaucomas - Primary open angle glaucoma - Ocular hypertension - Normal Tension Glaucoma - Primary angle closure glaucoma (Primary angle closure suspect, Intermittent glaucoma, acute congestive, chronic angle closure) - Secondary Glaucomas - Management : common medications, laser intervention and surgical techniques	
	Total hours	**92**

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6. DISPENSING OPTICS & MECHANICAL OPTICS CONTACT LENS & LOW VISION AIDS

DISPENSING OPTICS – MECHANICAL OPTICS

No.	Topic	No. of Lectures
1	Components of spectacle prescription & interpretation, transposition, Add and near power relation	1
2	Frame selection – based on spectacle prescription, professional requirements, age group, face shape	4
3	Measuring Inter-pupillary distance (IPD) for distance & near, bifocal height	1
4	Lens & Frame markings, Pupillary centers, bifocal heights, Progressive markings & adjustments – facial wrap, pantoscopic tilt	1
5	Recording and ordering of lenses (power, add, diameter, base, material, type, lens enhancements)	1
6	Neutralization – Hand & lensometer, axis marking, prism marking	3
7	Faults in spectacles (lens fitting, frame fitting, patients complaints, description, detection and correction)	2
8	Final checking & dispensing of spectacles to customers, counseling on wearing & maintaining of spectacles, Accessories – Bands, chains, boxes, slevets, cleaners, screwdriver kit	2
9	Spectacle repairs – tools, methods, soldering, riveting, frame adjustments	1
10	Special types of spectacle frames • Monocles • Ptosis crutches • Industrial safety glasses • Welding glasses	1
12	Frame availability in Indian market	2
13	FAQ's by customers and their ideal answers	2
	Total number of Hours	**21**

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CONTACT LENS

No.	*Topics*	No of Lecture(s)
1.	Introduction to Contact lenses ➢ Definition ➢ Classification / Types	1
2.	History of Contact Lenses	1
3.	Optics of Contact Lenses ➢ Magnification & Visual field ➢ Accommodation & Convergence ➢ Back & Front Vertex Power / Vertex distance calculation	3
4.	Review of Anatomy & Physiology of ➢ Tear film ➢ Cornea ➢ Lids & Conjunctiva	2
5.	Introduction to CL materials ➢ Monomers, Polymers	2
6.	Properties of CL materials ➢ Physiological (Dk, Ionicity, Water content) ➢ Physical (Elasticity, Tensile strength, Rigidity) ➢ Optical (Transmission, Refractive index)	3
7.	Indications and contraindications	2
8.	Parameters / Designs of Contact Lenses & Terminology	3
9.	RGP Contact Lens materials	1
10.	Manufacturing Rigid and Soft Contact Lenses – various methods	1
11.	Pre-Fitting examination – steps, significance, recording of results	3
12.	Correction of Astigmatism with RGP lens	2

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13.	Types of fit – Steep, Flat, Optimum – on spherical cornea with spherical lenses	1
14.	Types of fit – Steep, Flat, Optimum – on Toric cornea with spherical lenses	1
15.	Calculation and finalising Contact lens parameters	1
16.	Ordering Rigid Contact Lenses – writing a prescription to the Laboratory	1
17.	Checking and verifying Contact lenses from Laboratory	1
18.	Modifications possible with Rigid lenses	1
19.	Common Handling Instructions ➢ Insertion & Removal Techniques ➢ Do's and Dont's	1
20.	Care and Maintenance of Rigid lenses ➢ Cleaning agents & Importance ➢ Rinsing agents & Importance ➢ Disinfecting agents & importance ➢ Lubricating & Enzymatic cleaners	3
21.	Follow up visit examination	1
22.	Complications of RGP lenses	2
23.	SCL Materials & Review of manufacturing techniques	2
24.	Comparison of RGP vs. SCL	1
25.	Pre-fitting considerations for SCL	2
26.	Fitting philosophies for SCL	1
27.	SCL fitting assessment	2
28.	Types of fit – Steep, Flat, Optimum	3
29.	Calculation and finalising SCL parameters	2
30.	Disposable lenses a) Advantages and availability	1
31.	Soft Toric CL ➢ Stabilization techniques ➢ Parameter selection	2

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	➢ Fitting assessment	
32.	Common Handling Instructions ➢ Insertion & Removal Techniques ➢ Do's and Dont's	1
33.	Care and Maintenance of Soft lenses ➢ Cleaning agents & Importance ➢ Rinsing agents & Importance ➢ Disinfecting agents & importance ➢ Lubricating & Enzymatic cleaners	2
34.	Follow up visit examination	2
35.	Complications of Soft lenses	4
36.	Therapeutic contact lenses ➢ Indications ➢ Fitting consideration	1
37.	Specialty fitting ➢ Aphakia ➢ Pediatric ➢ Post refractive surgery	3
38.	Introduction to Bifocal CL	1
	Total Number of lectures	**67**

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LOW VISION AID

	Topics	Number of lectures
1	Definitions & classification of Low vision	1
2	Epidemiology of low vision Model of low vision service	1
3	Pre-clinical evaluation of low vision patients – prognostic & psychological factors; psycho-social impact of low vision	1
4	Types of low vision aids – optical aids, non-optical aids & electronic devices	3
5	Optics of low vision aids	1
6	Clinical evaluation – assessment of visual acuity, visual field, selection of low vision aids, instruction & training	3
7	Pediatric Low Vision care	4
8	Low vision aids – dispensing & prescribing aspects	1
9	Visual rehabilitation & counseling	1
10	Legal aspects of Low vision in India	1
11	Case Analysis	5
	Total hours	**21**

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7. BINOCULAR VISION & SQUINT

No.	Name of the topic	Number of lectures
1.	Binocular Vision and Space perception. ➢ Relative subjective visual direction. ➢ Retino motor value ➢ Grades of BSV ➢ SMP and Cyclopean Eye ➢ Correspondence, ➢ Fusion, Diplopia, Retinal rivalry ➢ Horopter ➢ Physiological Diplopia and Suppression ➢ Stereopsis, Panum's area, BSV. ➢ Stereopsis and monocular clues - significance. ➢ Egocentric location, clinical applications. ➢ Theories of Binocular vision.	6
2.	Anatomy of Extra Ocular Muscles. ➢ Rectii and Obliques, LPS. ➢ Innervation & Blood Supply. Physiology of Ocular movements. ➢ Center of rotation, Axes of Fick. ➢ Action of individual muscle. Laws of ocular motility ➢ Donder's and Listing's law ➢ Sherrington's law ➢ Hering's law Uniocular & Binocular movements - fixation, saccadic & pursuits. ➢ Version & Vergence. ➢ Fixation & field of fixation	4

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3.	Near Vision Complex Accommodation - Definition and mechanism (process). - Methods of measurement. - Stimulus and innervation. - Types of accommodation. - Anomalies of accommodation – aetiology and management.	3
4.	Convergence - Definition and mechanism. - Methods of measurement. - Types and components of convergence - Tonic, accommodative, fusional, proximal. - Anomalies of Convergence – aetiology and management.	5
5.	Sensory adaptations Confusion	1
6.	Suppression Investigations Management Blind spot syndrome	4
7.	Abnormal Retinal Correspondence Investigation and management Blind spot syndrome	1
8.	Eccentric Fixation Investigation and management	1
9.	Amblyopia Classification Aeitiology Investigation Management	4
10	Neuro-muscular anomalies Classification and etiological factors	1
11	History – recording and significance.	1
12	Convergent strabismus - Accommodative convergent squint - Classification - Investigation and Management - B Non accommodative Convergent squint - Classification	4

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	➢ Investigation and Management	
13	Divergent Strabismus Classification A& V phenomenon Investigation and Management	3
14	Vertical strabismus Classification Investigation and Management	1
15	Paralytic Strabismus Acquired and Congenital Clinical Characteristics Distinction from comitant and restrictive Squint	3
16	Investigations ➢ History and symptoms ➢ Head Posture ➢ Diplopia Charting ➢ Hess chart ➢ PBCT ➢ Nine directions ➢ Binocular field of vision	12
17	Non surgical Management of Squint	2
18	Restrictive Strabismus Features ➢ Musculo fascical anomalies ➢ Duane's Retraction syndrome ➢ Clinical features and management ➢ Brown's Superior oblique sheath syndrome ➢ Strabismus fixus ➢ Congenital muscle fibrosis	3

19	Surgical management	1
	Total Number of Hours	**60**

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8. COMMUNITY OPTOMETRY

Unit 1

PAEDIATRIC, GERIATRIC OPTOMETRY

No	Topics	Number of Lectures
1	The Development of Eye and Vision	2
2	History taking Paediatric subjects	2
3	Assessment of visual acuity	1
4	Normal appearance, pathology and structural anomalies of a) Orbit, Eye lids, Lacrimal system,	2
	b) Conjunctiva, Cornea, Sclera Anterior chamber, Uveal tract, Pupil	2
	c) Lens, vitreous, Fundus	1

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	Oculomotor system	
5	Refractive Examination	2
6	Determining binocular status	1
7	Determining sensory motor adaptability	1
8	Compensatory treatment and remedial therapy for : Myopia, Pseudomyopia, Hyperopia, Astigmatism, Anisometropia, Amblyopia	2
9	Remedial and Compensatory treatment of Strabismus and Nystagmus	2
10	Paediatric eye disorders : Cataract, Retinopathy of Prematurity, Retinoblastoma, Neuromuscular conditions (myotonic dystrophy, mitochondrial cytopathy), and Genetics	3
11	Anterior segment dysgenesis, Aniridia, Microphthalmos, Coloboma, Albinism	2
12	Spectacle dispensing for children	3
13	Paediatric contact lenses	2
14	Low vision assessment in children	2
	Total Number of Lectures	**30**

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GERIATRIC OPTOMETRY

No.	Topics	Number of Lectures
1	Structural , and morphological changes of eye in elderly	2
2	Physiological changes in eye in the course of aging.	2
3	Introduction to geriatric medicine – epidemiology , need for optometry care, systemic diseases (Hypertension, Atherosclerosis, coronary heart disease, congestive Heart failure, Cerebrovascular disease, Diabetes, COPD)	3
4	Optometric Examination of the Older Adult	2
5	Ocular diseases common in old eye, with special reference to cataract, glaucoma, macular disorders, vascular diseases of the eye	4
6	Contact lenses in elderly	1
7	Pharmacological aspects of aging	2
8	Low vision causes, management and rehabilitation in geriatrics.	4
9	Spectacle dispensing in elderly – Considerations of spectacle lenses and frames	4
	Total Number of Lectures	25

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Semester	Procedures	Minimum Number	Comments
IIyear	Accommodation	(1 case in presbyopic age)	
	Pushup test (Near point of Convergence)	10cases	
	Stereopsis test	10cases	
	Tear Breakup time	10cases	
	Amsler's Grid test	10cases (simulate)	Simulation of various conditions
	Color vision test	10cases	
	Schirmer's test	10cases	
	Confrontation test	10cases	
	Slit lamp illumination	3cases	
	Slit lamp examination	10cases	
	Finger tension	10cases (Normals)	
	Schiotz Tonometry	10cases (Normals)	
	Applanation Tonometry	10 cases (Normals)	
	Negative Relative Accommodation	10 cases	
	Positive Relative Accommodation	10 cases	
	Von Herick Grading of Anterior chamber depth	10 cases	
	Accommodative facility(±2.00D)	10cases	
	IPD	10cases	
	Proptosis evaluation	1demo	Video demonstration of cases
	Ptosis evaluation	1demo	Video demonstration of cases
	Pupillary evaluation -Direct -Consensual -RAPD	10cases	
	HVID	10cases	
	Maddoxrod (Phoria)	10cases	
	Negative Fusional vergence	10cases	
	Positive Fusional Vergence	10cases	
	Retinoscopy- Static, Dynamic and Cycloplegic Retinoscopy	25+25+25 cases	Model eye for retinoscopy
	Keratometry	25cases	
	Subjective Refraction JCC Clock Dial Duochrome Borish Delayed	25cases	
	Addition calculation	25cases	Give more simulated problems And discuss on it

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	Direct ophthalmoscope	10cases (Normals)	Show slides of various commonly seen retinal conditions
	Visual Field chart interpretation	10cases– discussion	Both kinetic and Static
	Bscan Interpretation	10cases– discussion	Discuss various abnormal cases
	Ascan chart Interpretation	10cases– discussion	Discussion having different Types of wave patterns
	Case Analysis	10cases	
	+90Dlens	10cases (Normals)	Slides of various Cup :Disc Ratios can be shown
III year	Gonioscopy	5cases (Normals)	Slides of abnormal angles
	Posting in optometry clinics	5+5+5+5+10 cases	Pediatric /contact lens/Low vision/ Orthoptics/ GOPD
	Camps	4camps	School screening, Cataract
	IDO(one achother)	10 Cases (Normals)	Slides of abnormal fundus
	Case Analysis -Pathology -Binocular Vision -Clinical Refraction -Dispensing optics	5+5+ 5+ 5 cases	
IV year **CLINICAL INTERNSHIP**	General OPD (History taking–DO)	500cases	Case reports should be submitted at the end of the postings
	Contact Lens	20cases(5 RGP+5Soft +5 toric)	Case reports should be submitted at the end of the postings
	Opticals	100cases	Case reports should be submitted at the end of the postings
	Low Vision care Clinic	10cases	Case reports should be Submitted at the end of the postings
	Binocular Vision clinic	10cases	Case reports should be submitted at the end of the postings
	Ophthalmology clinic (Common eye conditions)	50cases	Case reports should be submitted at the end of the postings
	Camps	10camps	Camp report submission

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BACHELOR OF SCIENCE IN PERFUSION TECHNOLOGY

Course duration

Duration shall be for a period of four years including one year internship training

Clinical activity report

- Practicals
- Clinical posting in cardiovascular department
- Peripheral posting in various departments- Echo, Cath lab, X- ray, blood bank and central lab

Practical examination

- Two practical examinations at the end second year
  - Applied pathology and applied microbiology - combined
  - Introduction to perfusion technology
- Three practical examinations at the end of the third year
  - Perfusion Technology – Clinical
  - Perfusion Technology – Applied
  - Perfusion Technology – Advanced

Internship

- Eligibility for internship Only after passing the 3rd year examination the student will peruse internship.
- The internship will consist of compulsory rotating practical training in the various subjects
- Duration one year each student should undergo one month in Paediatric Surgery, One month in thoracic surgery and one month in vascular surgery.
- If any of these specialty is not available in the hospital where the candidate studied he/she may be sent to the nearest reputed center in Kerala for training.

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Distribution of teaching hours in first year subjects

SL NO	Subject	Theory No Of Hrs	Practical No Of Hrs	Total No. of Hours
1	Human Anatomy	70	20	90
2	Physiology	70	20	90
3	Biochemistry	70	20	90
4	Pathology-[Clinical pathology, Haemotology & Blood –Banking]	70	20	90
5	Microbiology	70	20	90
	Total	350	100	450

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Distribution of teaching hours in second year subjects

SL NO	Subject	Theory	Practical	Clinical Posting	Total Hours
1	Medicine relevant to	50	-	-	50
2	Section A Applied Pathology	30	30	-	60
3	Section B Applied Microbiology	30	30		60
4	Applied Pharmacology	50	-		50
5	Introduction to Perfusion Technology	80	100	650	830
	Total	240	160	65	1050

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Distribution of teaching hours in second year subjects

SL No	Subjec	Theory	Practical	Clinical posting	Tota l
1	Perfusion Technology – Clinical	50	50	250	350
2	Perfusion Technology – Applied	50	50	25	350
3	Perfusion Technology –	50	50	25	350
	Total	150	150	750	1050

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Participant list

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6	ATHIRA.P.S
7	DHANYA SUNNY
8	FABIDA.P.V
9	FATHIMA
10	GANIYA ADHAM
11	HARITHA.P.C
12	INSHADA.E
13	JIFNA.PP
14	KEERTHANA.B
15	NASLIYATH.A.P
16	NEETHU BENY.T.M
17	RESA PAULSON
18	SANJAY.T.S
19	SARITHA.T
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21	SHAHANA.P.K
22	SHAMLA.K
23	SHEHEENA.K.P
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18	SHIFASHERIN.VP
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10	FASEELA.P.C
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18	MOHAMMED ASLAM
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18	PRAVEENA T S
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16	LIYA FATHIM
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12	JASLA SHERIN.A
13	JURIYA M.K.
14	LEELA.J
15	LULU M
16	MUBARAK.M
17	MUHAMMED NIYAS.K.K
18	MUHAMMED SHAMEEM.V
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24	SHAHANA
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27	SHIFANA.D
28	SILPA.K.V
29	UNAIS.K.V
30	VIBISHA.K

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CPT - 2019 Batch

SL No	Name
1	FATHIMATH NUSRIN
2	FAYISA SADEEM
3	HUDHA ABDUL JALEEL

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SL No	Name
1	ABHINAV V
2	ADHIYA FAIZAL
3	ADITHYA K.P
4	AJMAL ROSHAN.C
5	AKHIL S
6	ANJANA K P
7	ATHIRA P
8	AYSHA
9	BILAL.N
10	DHANUSHA.V.P
11	FATHIMA HIBA
12	FIDA V
13	HAIFA. AK
14	HANEENA MAHROOF O A
15	JAFER SADIQUE P T
16	JOHN JOSEPH
17	JYOTHY KRISHNA C B
18	MRUTHULA P
19	MUHAMED ASHFAQ
20	MUHAMMAD MUBAYIS
21	RINISHA DAS. T. P
22	RIZWIN AZEEZ
23	SANDRA. S
24	SHAHLA.N.P
25	SHIFA NARGEES
26	SHIHANA NASRIN.P.S
27	SNEHA. S. A
28	SNEHITH VISWAS V
29	SOLJA T V
30	SUHAIL C

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27	SNEHA. S. A
28	SNEHITH VISWAS V
29	SOLJA T V
30	SUHAIL C

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Sl. No	Name
1	AISWARYA T
2	ANJANA P PRAKASH
3	ANUSHA P
4	ARYA T
5	ASHIQUALI P
6	ASNA K
7	ATHIRA.K.K
8	AYSHA LIYA
9	FATHIMA FARHANA
10	FATHIMA NAFLA P K
11	FATHIMA SANA. V
12	FIDHA PARVIN
13	GAYATHRI V S
14	HANA FATHIMA
15	HARSHA. P. P
16	HASNA SHERIN.P
17	JAFINA MOIDU
18	KARTHIKA.S.S
19	MUHAMMED AL AMEEN. K. T
20	MUHAMMED NISAM V K
21	NAJIYA NASREEN. P
22	NASHWA MARYAM
23	NASNA. T. P
24	NIDHA FATHIMA E M K
25	NOURIN C
26	SHEETHAL M
27	SHINFA ASMI. P
28	VAISHNAVI. P. P
29	AISWARYA T

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SL No	Name
1	AFRIN K
2	AYSHA HANAN
3	AYSHATH RASHA MOHAMMED
4	DANIYA.KV
5	FATHIMA
6	FATHIMA K
7	FIDHA. K
8	HIBA FATHIMA
9	HIBA NUSRIN
10	HISHANA FATHIMA P.T
11	JABIR
12	LIYA FATHIMA P C
13	LIYA LATHEEF
14	MINNU C SAJI
15	MUHAMMED HASIL
16	MUHAMMED IRSHAD K P
17	MUHAMMED RASHID K A
18	NAJIYA ABOOBACKER
19	NANDANA SADANANDAN
20	NIDHA FATHIMA. M.P
21	NILOOFAR SULTHANA
22	RISTHANA M
23	SARANYA SAJEEV
24	SHABLA K P
25	SHAMILA M M
26	SHIBLA NARGEES . E
27	SREELAKSHMI P S
28	THANSILA RAHIMAN
29	THANVEERA JAHAN

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SL No	Name
1	ADHITH VINOD
2	ADHITHYA TOMY
3	AFNAN PUTHIYACHIRAKAL
4	AMAL SIDAN
5	AMEENA LULU K
6	ANANTHU C B
7	ARUN G KRISHNA
8	ATHILA NIJAMUDEEN
9	BINSON .B
10	DHEEMA SALAM
11	DILNA V K
12	DILNAWAS VETTASSERI
13	DILSHA .K.C
14	FATHIMA MIRSHA
15	FATHIMATHU SHAHANA
16	FIDHA THASNEEM.E.K
17	HIBA FATHIMA
18	HIBA THESNIEM . C
19	JAIHAJYOTHY P.K
20	KAVYA V.S
21	MARJANA THASLI
22	MISBA.K.
23	NADHA M
24	NAHLA SARIN. E
25	NIMISHA N
26	SHABEEB AHAMMED. K
27	SHADHIN.TK
28	SHAHANA SHIRIN. C
29	SHIYANA NASRIN
30	SREELAKSHMI A

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SL No	Name
1	DEVASENA S
2	FAHIDA K
3	GANGA K.V.
4	HIBA HAKEEM
5	RAHAFA.P.A

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SL No	Name
1	ABILA RAHMAN.K.T
2	ANGEL MARY
3	ASHNA ASHRAF
4	ASWANI.B
5	AYISHA LIYA
6	DEVIKA D
7	FAHEEMA RIYAS
8	FASNA N P
9	FATHIMA FIZA N V
10	FATHIMA NIHALA P T
11	FATHIMATH RANA V
12	FATHIMATH SUHARA
13	FIDA V
14	HANNA PARVEEN V K
15	HARSHA.T.V
16	HASNATH M K
17	JALVA JASEER
18	MALAVIKA S
19	MUHAMMED ANSIF C P
20	NAMITHA MARIA JIJI
21	NANDAGOPAL VIJAYAN
22	NASREEN
23	NIHALA. T
24	NIHALA V P
25	NIYA MARY JOSE
26	RENNA SIYAN T K
27	RIDA HANEEF. P
28	SAFEENA MK
29	SHYMA P
30	SIMI A
31	SONU P C

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SL No	Name
1	AFEEFA FARSANA
2	AFTAB AHAMMED J
3	ALFIYA V K
4	AMRITHA.K
5	ANJANA G R
6	ASWIN JABY JOHN
7	ATHULYA.C
8	AYSHA NIDHA
9	BINDHUJA B
10	BUNIYA SHAROOBI MT
11	DILSHA P P
12	DINSHA SALAM M
13	FATHIMA RINSHA
14	FATHIMA SAFA.K
15	FATHIN SHAMEEM
16	FIDA
17	JALWA P
18	LINSHA FATHIMA
19	MAJITHA FARSANA.K
20	MIDHUN MADHAVAN K
21	NABEELA NOUSHAR.A
22	NADA C P
23	NADIYA SHERIN
24	RAHEESA.K.M
25	RIYA.P
26	SAFEEDA.M
27	SHAHANA.K.V
28	SHANA SIRIN N P
29	SHASNA MOHAMMAD
30	SUSRUTHA V S

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SL No	Name
1	AGHINA M
2	AISWARYA R
3	ALEENA K R
4	ALEENA MAHAROOF M
5	ANAMIKA V S
6	ANCY A S
7	ANNMARIYA STUVERT
8	ATHIRA T A
9	AYSHA HANNA
10	FATHIMA DILNA P K
11	FATHIMA HUSNA T
12	FATHIMA SHAHANA
13	FATHIMA SHIFNA T P
14	HIBA KHADEEJA
15	KRISHNA PRAKASH C
16	MAZINA P V
17	MIRSANA AYISHA ISMAIL K K
18	MOHAMMED HIBATHULLA. E.K
19	MUBASHIRA K T
20	MUHAMMED SHAFEEQUE RAHMAN.V
21	NAHLA SHERIN.K.K
22	NANDANA N S
23	NEHA KAMATH
24	RANIYA FATHIMA.M K
25	RINSHA FEBIN K
26	SALIHA P
27	SANIA K
28	SHAHLA SHERIN
29	SHIFNA T
30	SUHA MUHAMMADALI

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SL No	Name
1	BUSHRA BASHEER
2	FATHIMATH HIBA C
3	SULALA MUNNA N.K

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SL No	Name
1	ABHIRAMI K S
2	AKSA SANTHOSH
3	ASHA NAZRIN
4	AYSHA JINU
5	AYSHA SHAFNI
6	BINZA BASHEER
7	DEVIKA JITHESH
8	FAMITHA A M
9	FATHIMA DILNA T
10	FATHIMA HIBA AMEER
11	FATHIMA NAZER
12	FATHIMA NOOFI
13	GOPIKA SANTHOSH
14	HANNA FATHIMA T T
15	JASIYA MUNAVVIRA.K.M
16	KADEEJA ASMI V
17	KHADEEJA NASHA
18	LIJINA JIJI
19	MUBASHIRA P P
20	NAJA FATHIMA
21	NAJIYA SALAM
22	NANDANA.P.P
23	NEHA A K
24	NIYA MANOJKUMAR K.V
25	RITHU PARVEEN C K
26	SANIYA U T
27	SHAHNAS ABDUL SATHAR P P
28	SREELAYA S
29	SUBITHA SURESHKUMAR
30	SULEKHA
31	SWATHI K

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SL No	Name
1	AANY MARIA GEORGE
2	ABDULLAH YAZEEN
3	ABHIJAY P
4	ADELIN JANET ROLLAND
5	ADHILA ASHARAF
6	AFRA SHAN E P
7	ANIL KUMAR.K.C
8	AYISHA AMNA M
9	CHAITHRA GIREESH P V
10	FATHIMMA ABDUL LATHEEF M V
11	HANA FATHIMA
12	HIBA JASMIN A K
13	HIMA K
14	HUSNA M
15	JUMANA JUBIN.A
16	LAHNA SHERIN.N
17	MIRSHA ASHRAF T
18	MOHAMED RINSHAN N K
19	NAIFA FATHIMA C K
20	NAJA M K
21	NANDANA K R
22	NIDHA NAJMUL.P
23	RISWIN KP
24	SAHLA.K.T
25	SAJNA.S
26	SHAMIL K
27	SHANA
28	SHARBINA SHERIN.P
29	SIYANA SHARHA M
30	SREYA RATHEESH

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SL No	Name
1	AAMINA VALLOPRA
2	AFNAN K
3	ANAGHARAJ S
4	ANAN MEHBOOB
5	ANUPAM SHANKAR
6	ASHIK ALI C M
7	ASIYA FATHIMA A
8	DEVIKA S
9	FATHIMA HANNA P C
10	FATHIMA NASNIN P
11	FATHIMA NASRIYA U
12	FATHIMA RISHA P V
13	FATHIMA RUFAIDHA T
14	FAYIZA O P
15	FIDHA MUSTHAFA
16	HIBA NASRIN
17	IRSHANA TC
18	JIFNA SHARIN P
19	JUMANA T A
20	MISNA FIDHA
21	NAJLA
22	NANDANA V K
23	NASEEFA MARIYAM
24	NASHVA T P
25	NISWA JAMSHI C
26	NYLA FATHIMA K P
27	SANA K
28	SHIMRIN K
29	SNEHA J R
30	SWATHY LAKSHMI A

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SLNo	Name
1	ALFINA BEEVI
2	SHARON

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SL No	Name
1	AMINA BEEGAM A S
2	ANJANA A P
3	ANUSREE C
4	AVEENA DENNIES
5	AYISHA HANA
6	AYSHA LIYANA T S
7	DEVA PRIYA SURESH
8	DEVIKA SAJIKUMAR
9	DIYA FATHIMA M K
10	DIYA FATHIMA P C
11	FATHIMA NIHANA.V
12	FATHIMA SHAFA M P
13	HIBA ABDUL AZEEZ.V.M
14	HYFAAR
15	JASMIYA N K
16	MARIYAMBI P K
17	MEHNAS
18	MUHAMMED SAFVAN O P
19	NAFLA M K
20	NASMIYA NAZAR
21	NITHYA K K
22	RASHA SHAREEF
23	R C RILFA
24	RINSHANA P A
25	SAFA HANEEFA
26	SANUSHNAV S
27	SHELNA SHERIN U K
28	SNEHA SANTHOSH
29	SREENIDHI.S
30	VINEETH.C

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